Submissions for Capillary Run - Sequencing

1. Sequencing reactions must be carried out in 96-well plates from ABI (catalog # N8010560) or similar semi-skirted 200-300 ul thin walled PCR plate with the notch at A12.
2. All sequencing reactions must be done using ABI BigDye Terminator v3.1 chemistry and following ABI's protocols.
3. The sequencing reactions must be cleaned up, i.e. unincorporated dye terminators must be removed, prior to submission. The most common methods for clean-up are ethanol precipitation, magnetic beads, e.g. Agencourt CleanSEQ, or spin columns. The method you choose will depend on your preference and available equipment. For optimum instrument performance, indicate the clean up method used in the comment section of the iLab submission page.
4. Plates must be brought to the Facility after the final drying step of the clean-up. Cover plates with adhesive plate sealers (NOT parafilm, foil, or hard plastic).
5. An Electronic Instrument Sample Sheet must be prepared for each plate using the 3730SampleSheet_Template_Sequencing.txt available on the iLab submission page. Open the provided template in Excel, fill out as specified below, save as a tab delimited text file and upload to iLab.
   - Open template in Excel to complete
   - Enter "plate name" in cells A2 and B2
     - Name must be unique and concise
     - Name must match Sample Sheet filename
     - Match filename to the 96-well plate name for ease in identification
     - **NO periods, spaces, \ / : * ? " < > | or _**
   - Enter Sample IDs in Column B
     - Use only alphanumeric characters, underscore or dash
     - **NO periods, spaces, \ / : * ? " < > | or _**
   - Save as tab delimited text file (.txt)
   - Email completed Instrument Sample Sheet(s) to gpclseq@pitt.edu